

The Genome of the Kinetoplastid Parasite, *Leishmania major*

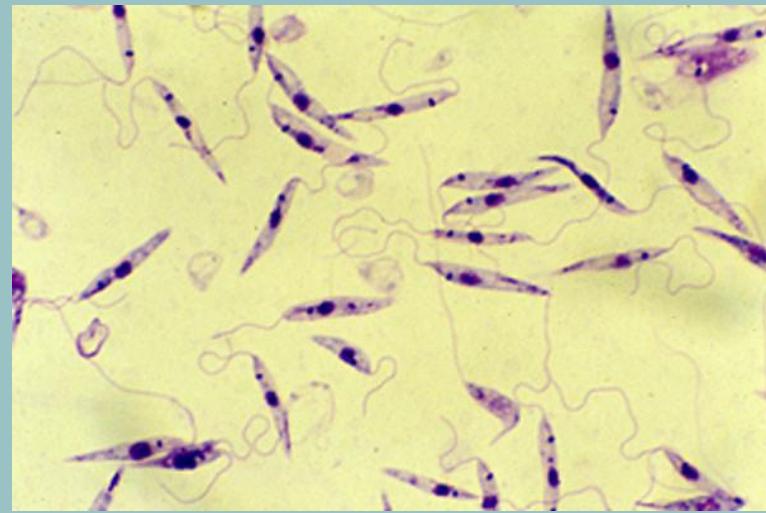
A Journal Club Presentation by
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Biological Databases
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Outline

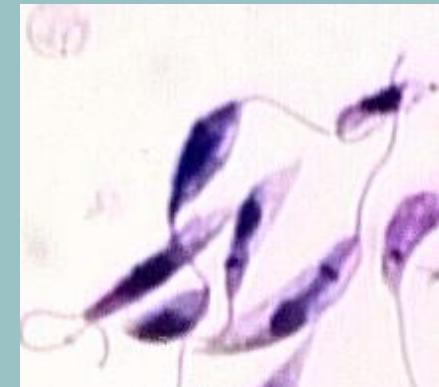
- Why Study Leishmania
- Research Methods
- Genome Structure and Contents
- Significant Genetic Findings



[http://www.uni-tuebingen.de/modeling/
Mod_Leish_Intro_en.html](http://www.uni-tuebingen.de/modeling/Mod_Leish_Intro_en.html)

Why Study Leishmania

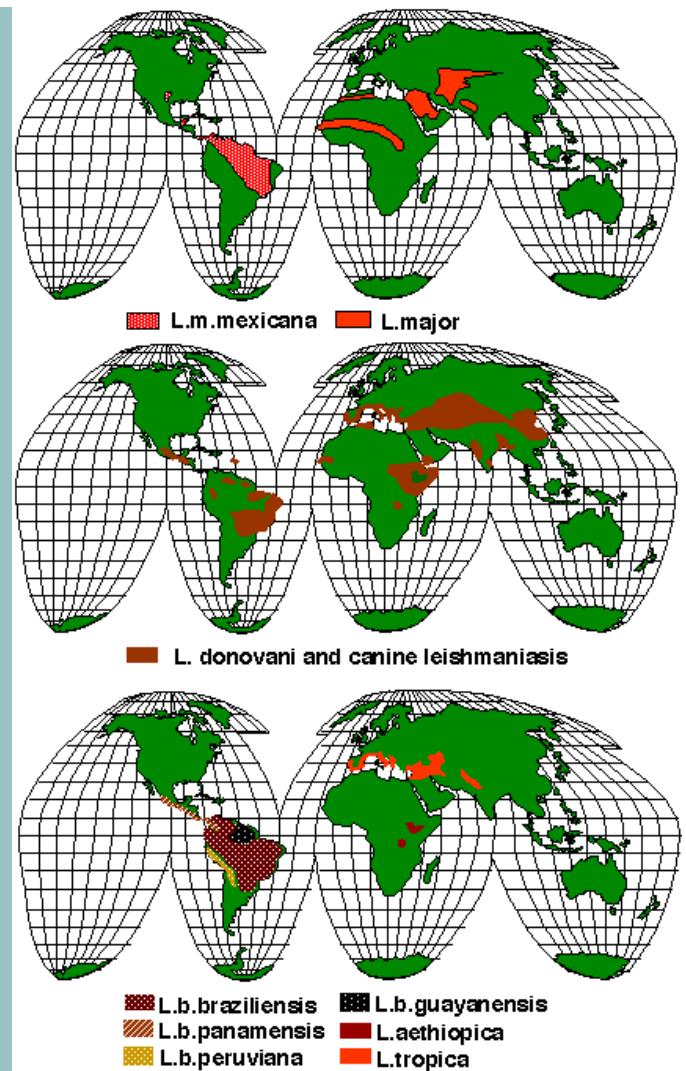
- Major pathogenic parasite
- 2 million infections in 88 countries annually
- spectrum of diseases: “leishmaniases”



[http://www.icp.ucl.ac.be/
~opperd/parasites/images/
promast.jpg](http://www.icp.ucl.ac.be/~opperd/parasites/images/promast.jpg)

Leishmania is uniquely successful parasite

- Tropic/sub-tropic
- Uniquely adapted to avoid host-destruction
- Thrive and proliferate due to unique glycoconjugates on outside of cells



Symptoms of the Leishmaniases

- Skin sores and ulcers
- Difficulty breathing/swallowing
- Eroding away in mouth, tougue, gums, lips, and nose

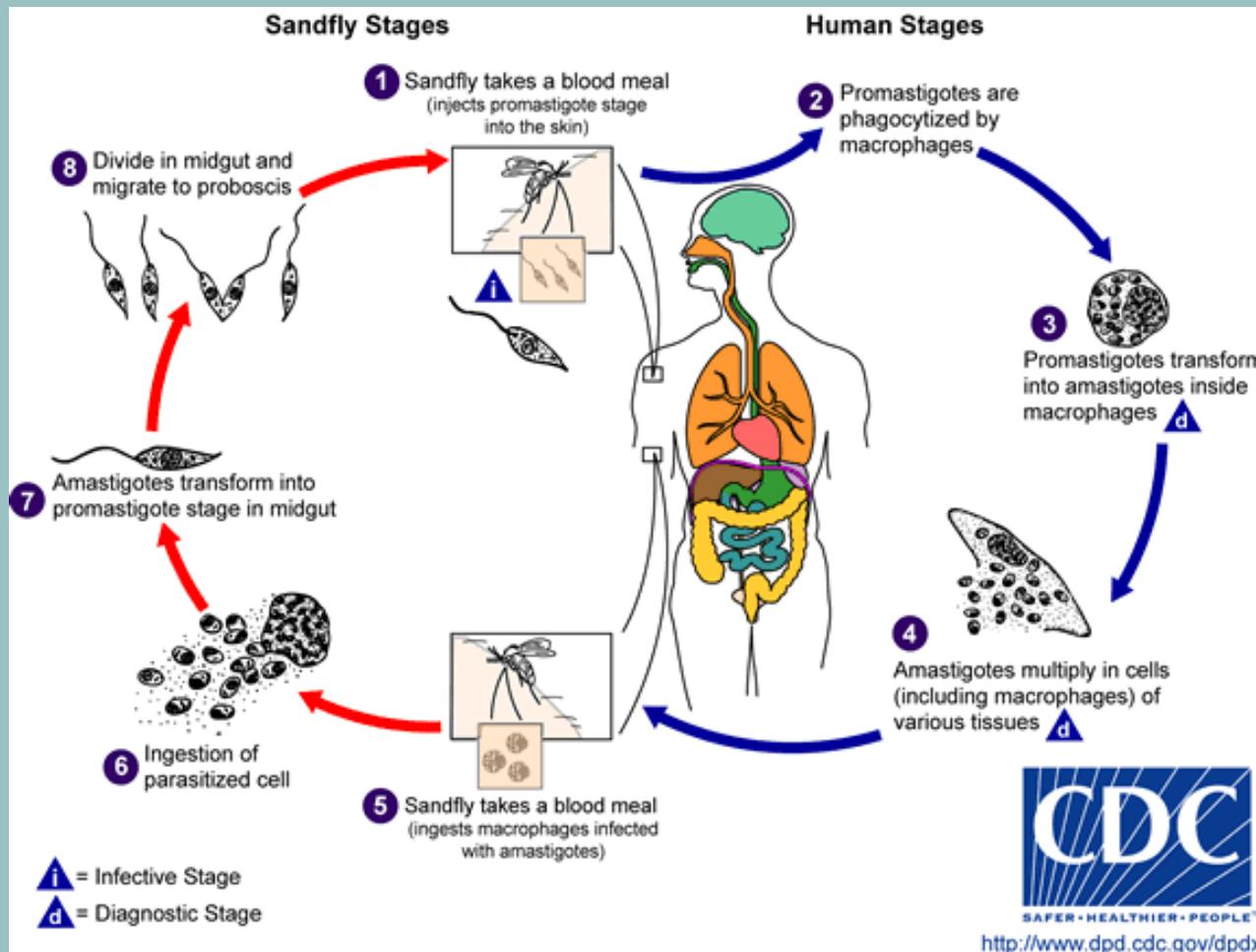


<http://anthropology.net/2009/09/10/ancient-leishmaniasis-from-coyo-oriente-cemetery-in-chile/>



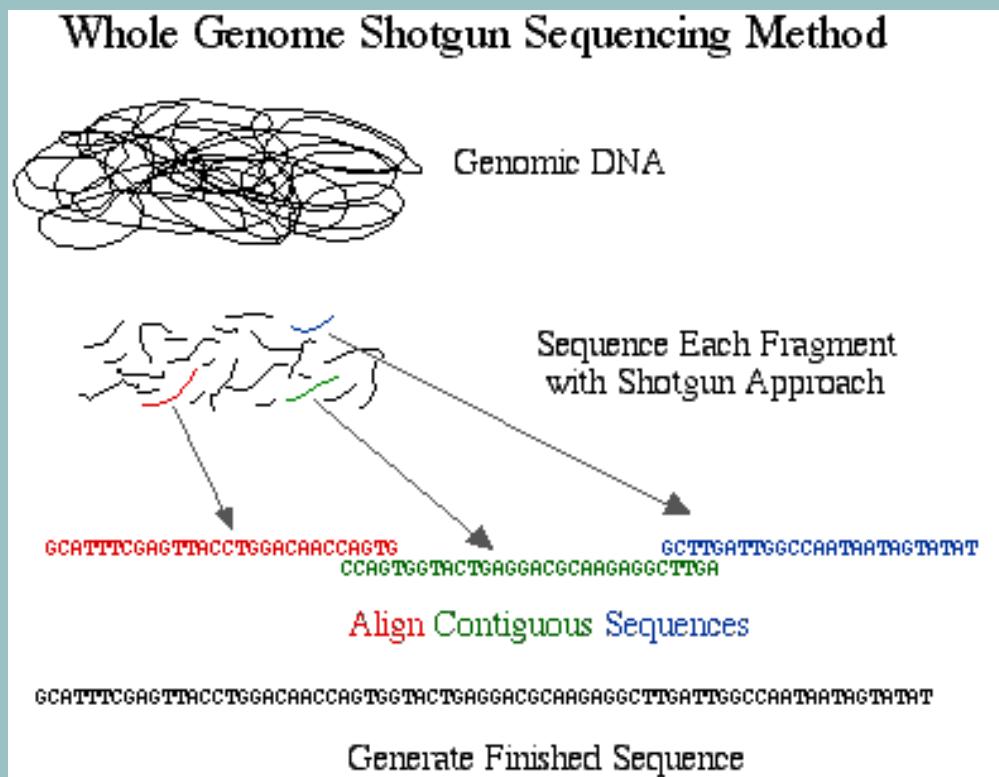
<http://adams Spencerphotography.com/leishman/>

Life cycle of Leishmania



Shotgun Sequencing

- DNA broken up into many segments
 - Sequenced to obtain *reads*
 - overlapping *reads* assembled to a continuous sequence
- Sequences compared between 3 trypanosomatids, “tritryps”



Leishmania major genome structure and content

- 32,816,678 base pairs
- 36 chromosomes
- Accuracy was compared to an optical map
- Smaller gene families
- Larger gene families

Parameter	Number
<i>The genome</i>	
Size (bp)	32,816,678
G+C content (%)	59.7
Chromosomes	36
Sequence contigs	36
Percent coding	47.9
<i>Protein-coding genes</i>	
Genes	8272
Pseudogenes	39
Mean CDS length (bp)	1901
Median CDS length (bp)	1407
G+C content (%)	62.5
Gene density (genes per Mb)	252
<i>Intergenic regions *</i>	
Mean length (bp)	2045
G+C content (%)	57.3
<i>RNA genes</i>	
tRNA	83
rRNA‡	63
siRNA‡	63
snRNA	6
snoRNA	695
srpRNA	1

* Region between protein-coding CDS.

‡ The exact number cannot be determined because of misassembly.

Table 1: Summary of L. major genome

Some genes are unique to *L. major* when compared to those in *T. brucei* and *T. cruzi*

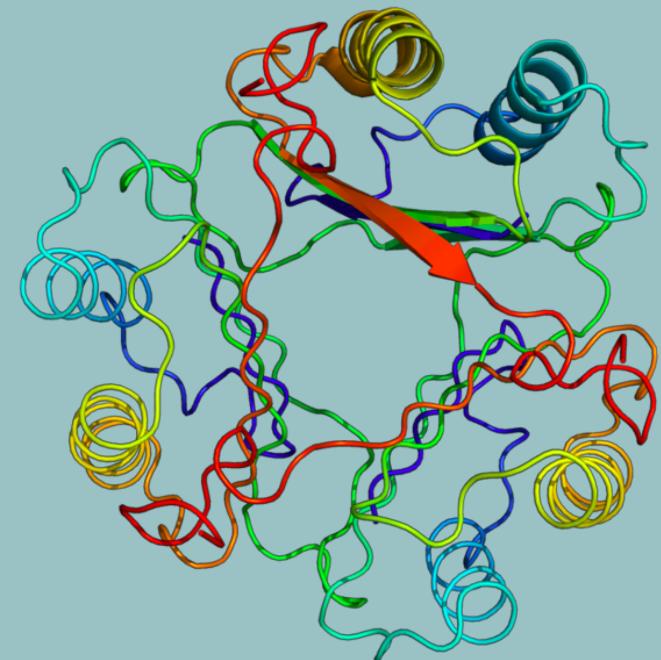
- *L. major* telomeres are distinct heterogeneous in structure
- 910 *L. major* genes have no orthologs in other 2 Tritryp genomes
- “*Leishmania* restricted genes”
 - Randomly distributed in genome
 - Some responsible for metabolic differences
 - 68% have unknown functions

Family size*	Gene product(s)	<i>L. major</i> —specific	Organization†	Chromosome(s)
491	Hypothetical proteins (several annotations)	Some	D	Multiple
189	Kinesins/hypothetical proteins	Some	T+D	Multiple
60	Protein kinases (several groups)	Some	T+D	Multiple
46	Amastins	Most	T+Tl+D	8, 31, 34, 36
32	Protein kinases (CMGC group)	One	D	Multiple
32	PSA-2 (GP46)	All	T+D	12, 21, 31, 35
29	RNA helicases/eIF-4a	None	T+D	Multiple
27	ATPase/serine peptidases	None	D	Multiple
29	Hypothetical proteins (kinesin-like)	One	D	Multiple
25	Protein phosphatases	None	T+D	Multiple
25	Tuzins	Some	Tl+D	8, 34, 36
24	Protein kinases (STE group)	Some	D	Multiple
23	Amino acid permeases	Some	T+D	Multiple
19	HSP83	None	T+D	29, 33
18	DNA helicases	Some	D	Multiple
18	β-tubulins	None	T+D	8, 21, 33
17	Hypothetical proteins (LACK)	One	D	Multiple
17	Hypothetical proteins	Some	T+D	11, 13, 21, 29, 31, 36
15	Calpain-like cysteine peptidases	Some	T+D	4, 20, 25, 31, 36
14	HSP70 and related proteins	None	T+D	1, 18, 26, 28, 30, 35
14	Phosphoglycan β 1,3 galactosyltransferases	Some	T+D	2, 7, 14, 21, 25, 31, 35, 36
14	Dynein heavy chain	One	D	Multiple
14	RNA helicases	None	D	Multiple
14	α,γ-ε-tubulins	Bone	T+D	13, 21, 25
13	Hypothetical proteins (PIP-like protein)	One	D	Multiple
13	Pteridine transporters	Some	T+D	4, 6, 10, 19, 35

Table 2: *L. major* Friedlin protein coding gene families

Two closely related genes of interest: LmjF33.1740 and LmjF33.1750

- Macrophage migration inhibition factor (MIF)
- MIF genes in humans encode a lymphokine protein mediator involved in innate immunity
- L. major MIFs vs. host MIFs
- May modulate host macrophage response to promote parasite survival



Structure of MIF protein
http://en.wikipedia.org/wiki/Macrophage_migration_inhibitory_factor

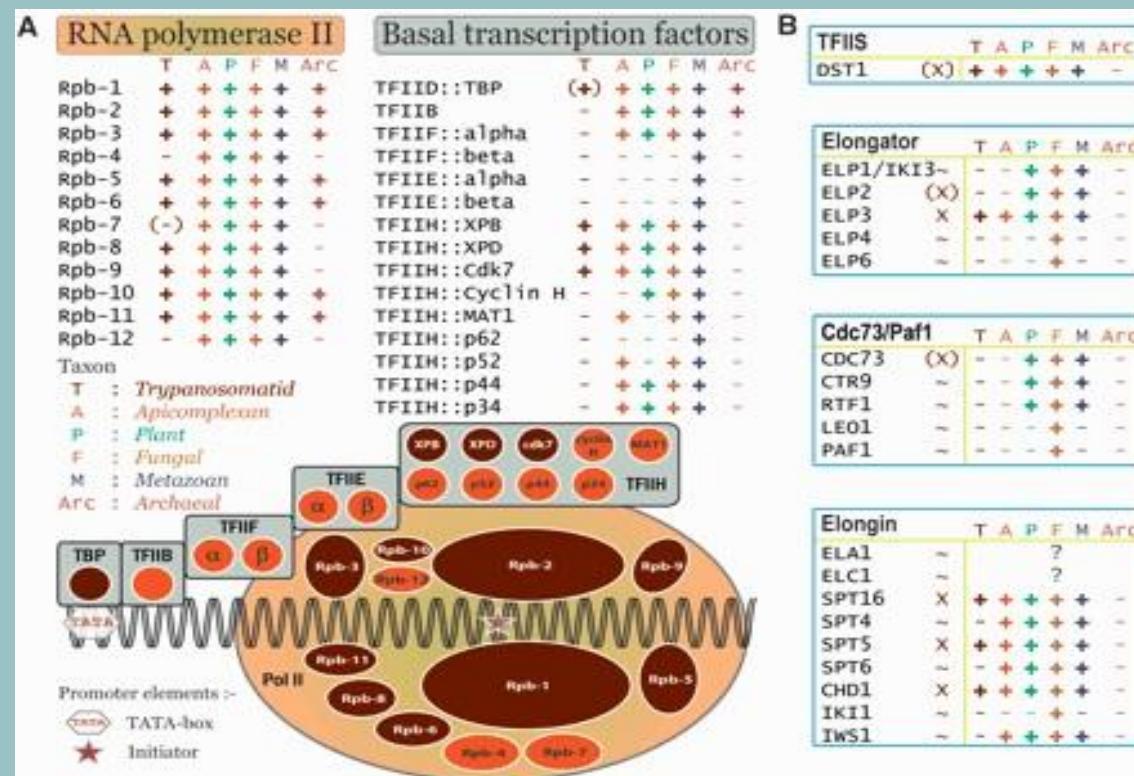
Significant Findings- Transcription Machinery of Leishmania Compared to Other Eukaryotes

Figure 1

RNA Polymerase II

- Protein Families
- Subunits
- Transcription

Factors



Significant Findings- Protein Families Involved in the Regulation of Gene Expression in Model Eukaryotes

Regulation of Gene Expression

Figure 2-
Gene Expression of Various
Protein Domains
by Species

		Lm	Tb	Tc
PF00642	349	42.3	17.7±4.7	Zinc finger C-x8-C-x5-C-x3-H type (and similar)
PF00097	349	114.7	62.6±32.0	Zinc finger, C3HC4 type (RING finger)
PF00096	78	307.6	136.6±100.8	Zinc finger, C2H2 type
PF00098	58	96.3	26.5±26.7	Zinc knuckle
PF01753	19	10.3	6.0±2.9	MYND finger
PF00856	15	22.5	13.6±3.7	SET domain
PF00533	11	2.4	8.9±4.0	BRCA1 C Terminus (BRCT) domain
PF00439	11	1.9	12.2±5.3	Bromodomain
PF02146	10	2.5	4.2±2.6	Sir2 family
PF00313	8	7.9	0.7	'Cold-shock' DNA-binding domain
PF00249	8	0.7	3.6±1.8	Myb-like DNA-binding domain
PF00643	7	104.4	24.3±20.7	B-box zinc finger
PF00412	6	2.0	18.9±14.0	LIM domain
PF00569	5	40.5	16.1±12.2	Zinc finger, ZZ type
PF02178	4	1.0	6.1±4.5	AT hook motif
PF01381	4	0.5	11.7	Helix-turn-helix
PF00628	4	0.5	8.0±2.5	PHD-finger
PF00505	4	2.9	0.9±0.6	HMG (high mobility group) box
PF00352	4	0.4	35.7	Transcription factor TFIID (or TATA-binding protein, TBP)
PF00382	3	0.4	23.1±9.0	Transcription factor TFIIB repeat
		1.0	41.6	1.2 [1] 1.1 [1] 1.0 [2]
		1.0	5.9	1.2 [1] 1.1 [1] 1.0 [2]
		0.7	1.5±1.1	1.2 [1] 1.1 [1] 0.5 [1]
		0.4	4.1	2.0±1.3

Significant Findings- Proteins Involved in Post Translational Modification are Potential Drug Targets

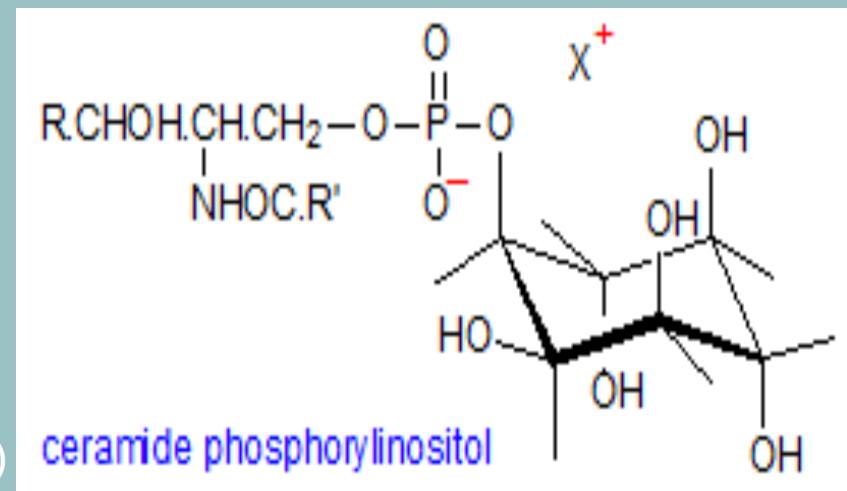
Post Translational Modification

- Protein modification typical of a eukaryote
 - Typical number of genes code for most protein modifications
- High number of genes code for myristylation and prenylation
- These enzymes could be drug targets due to high number of substrates

Significant Findings- Surface Molecules Unique to Leishmania are Potential Drug Targets

Surface molecules

- Surface molecules critical for pathogenic species
- Primary sphingolipid expressed is Inositol Phosphorylceramide (IPC)
- Not produced in mammals
- Enzymes that construct IPC are excellent drug targets given that this important surface molecule is not produced in humans



http://lipidlibrary.aocs.org/Lipids/glyP_ino/index.htm

Implications: Tritryp genomes help show unique biology of Leishmania and give insight to Eukaryote evolution

- Differences from other eukaryotes: post-translational modification
 - polycistronic gene clusters
 - mRNA trans-splicing coupled with polyadenylation
- Leishmania branched off from other eukaryotes very early on
 - Differences arose after branching off of Leishmania

Full genome provides crucial information for new therapies of Leishmaniasis

- analysis of virulence factors
- enzymes in metabolic pathways
- potential vaccine candidates



[http://www.dailymail.co.uk/health/
article-1191257/Ben-Fogle-I-nearly-lost-half-
face-flesh-eating-bug--The-cure-injections-
poison.html](http://www.dailymail.co.uk/health/article-1191257/Ben-Fogle-I-nearly-lost-half-face-flesh-eating-bug--The-cure-injections-poison.html)

Conclusion

- Leishmania: sub-tropical parasite that causes Leishmaniasis, an infection affecting 2 million people annually
- 36 chromosomes sequenced
 - compared to Tritryp organisms and humans to understand biology and function of Leishmania genes
- Drug targets were found in both post translational modification enzymes and in surface molecules
 - Full genome allows for analysis and development of therapies

Acknowledgments

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